

**GLIADIN BINDING TO CXCR3  
INDUCES A MyD88- AND G-COUPLED  
PROTEIN RECEPTOR-DEPENDENT  
INCREASED INTESTINAL  
PERMEABILITY AND ZONULIN  
RELEASE**

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# Background & premises

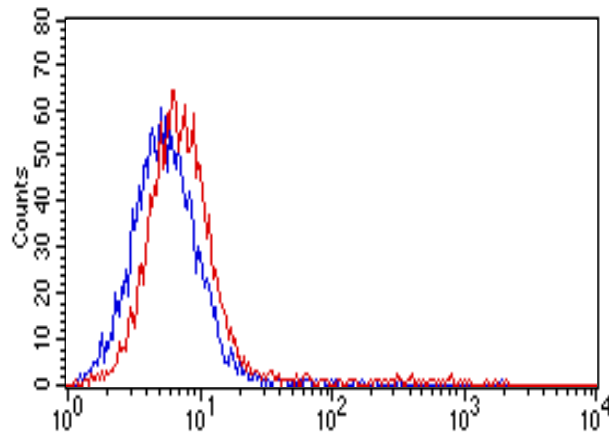
- Celiac disease (CD) is an auto-immune enteropathy triggered by the ingestion of gluten
- Gliadin, a component of the grain protein gluten, is known to induce **increased intestinal permeability**, which is considered an early crucial biological event in the pathogenesis of CD
- **Zonulin** induces tight junction disassembly. It is therefore considered to be involved in CD
- **In CD:**  
An increased and persistent release of zonulin and a significant increase in intestinal permeability (*S. Drago et al. Scand J Gastroenterol. 2006*)  
Apical, but not basolateral, exposure to gliadin led to zonulin release (*MG Clemente et al. Gut 2003*)
- We recently identified the chemokine receptor **CXCR3** as the receptor to which gliadin binds

# Aim

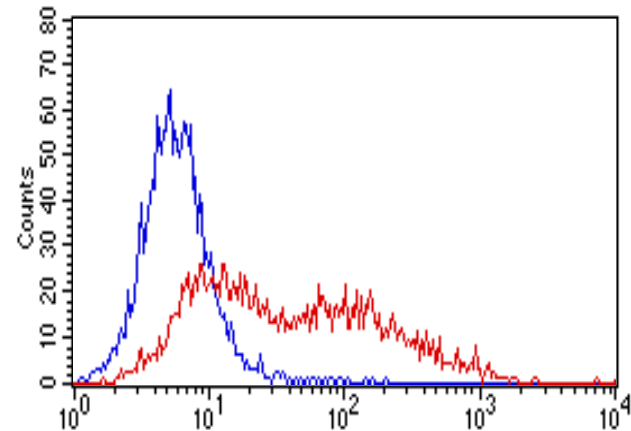
To explore the function of CXCR3 after gliadin binding

# CXCR3 expression on transfected HEK293T cells

pcDNA-transfected HEK cells



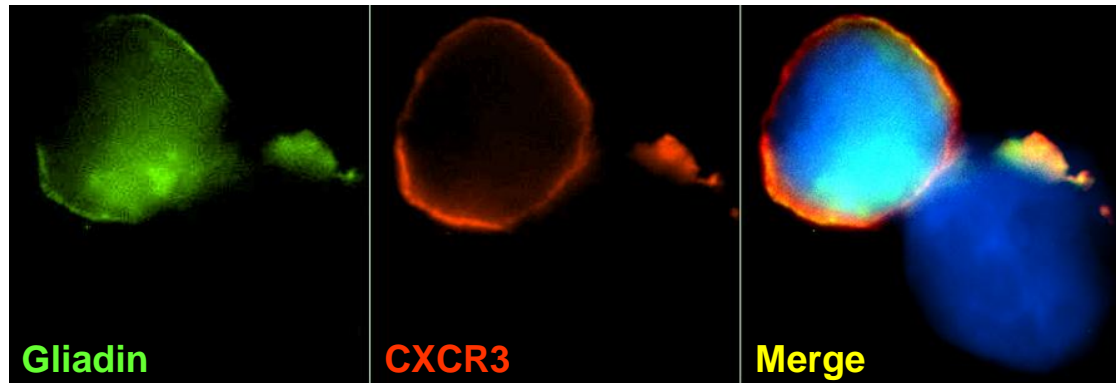
CXCR3-transfected HEK cells



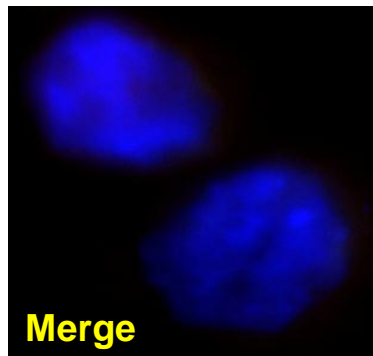
Isotype-matched control antibody (blue)  
Anti-CXCR3 antibody (red)

# Co-localization of PT-gliadin and CXCR3

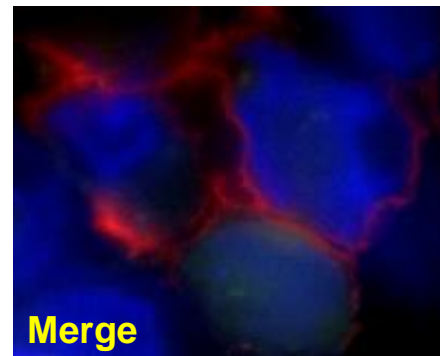
CXCR3-transfected HEK cells  
PT-gliadin treatment



pcDNA-transfected HEK cells  
PT-gliadin treatment

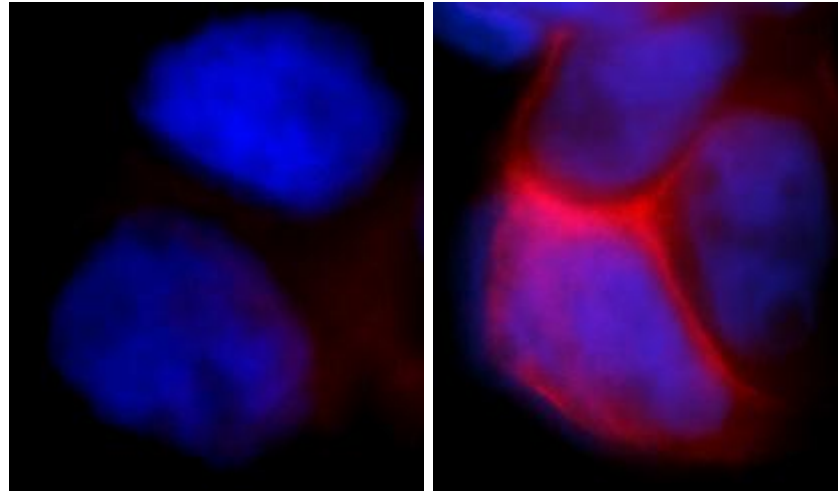


CXCR3-transfected HEK cells  
BSA treatment

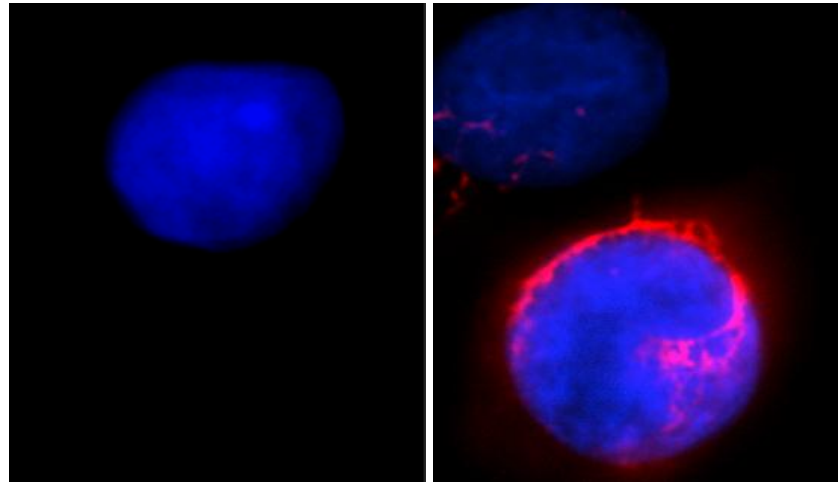


# Basal CXCR3 expression on intestinal epithelial cell lines

Caco-2



IEC6

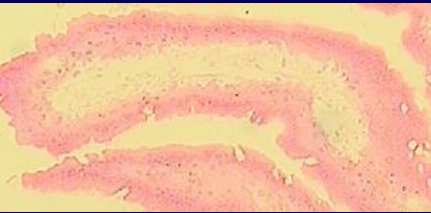
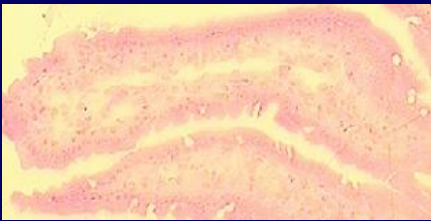


Isotype control

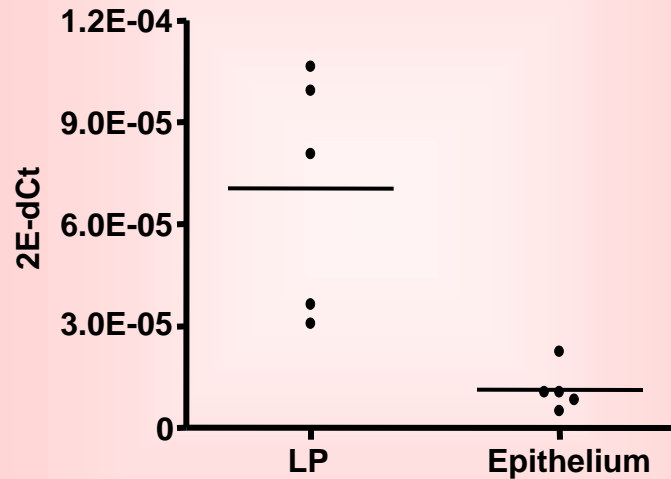
CXCR3

# CXCR3 is expressed in mouse intestinal epithelium and in lamina propria

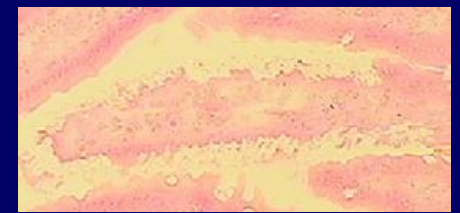
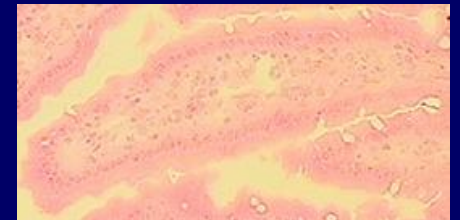
**Laser Capture  
Lamina propria preparation**



**Real Time RT-PCR  
CXCR3 expression**

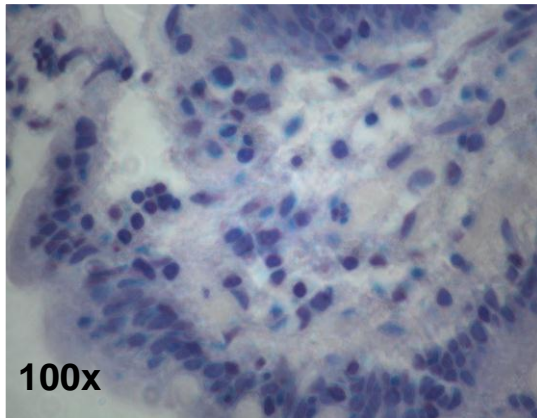
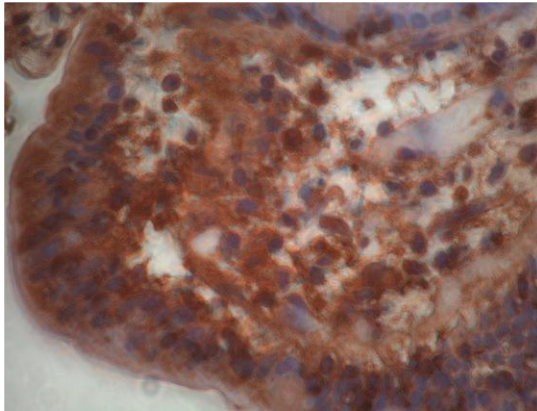


**Laser Capture  
Epithelial tissue preparation**

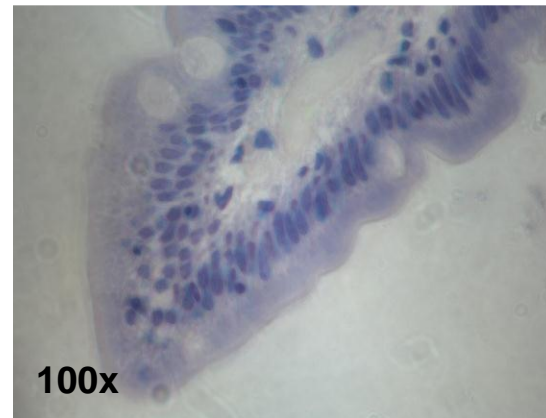
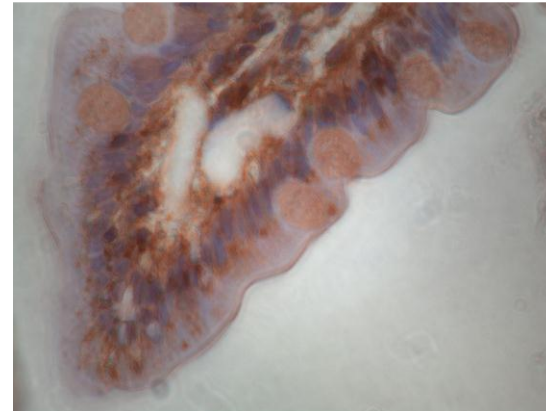


# CXCR3 expression in human intestinal tissues

Active CD

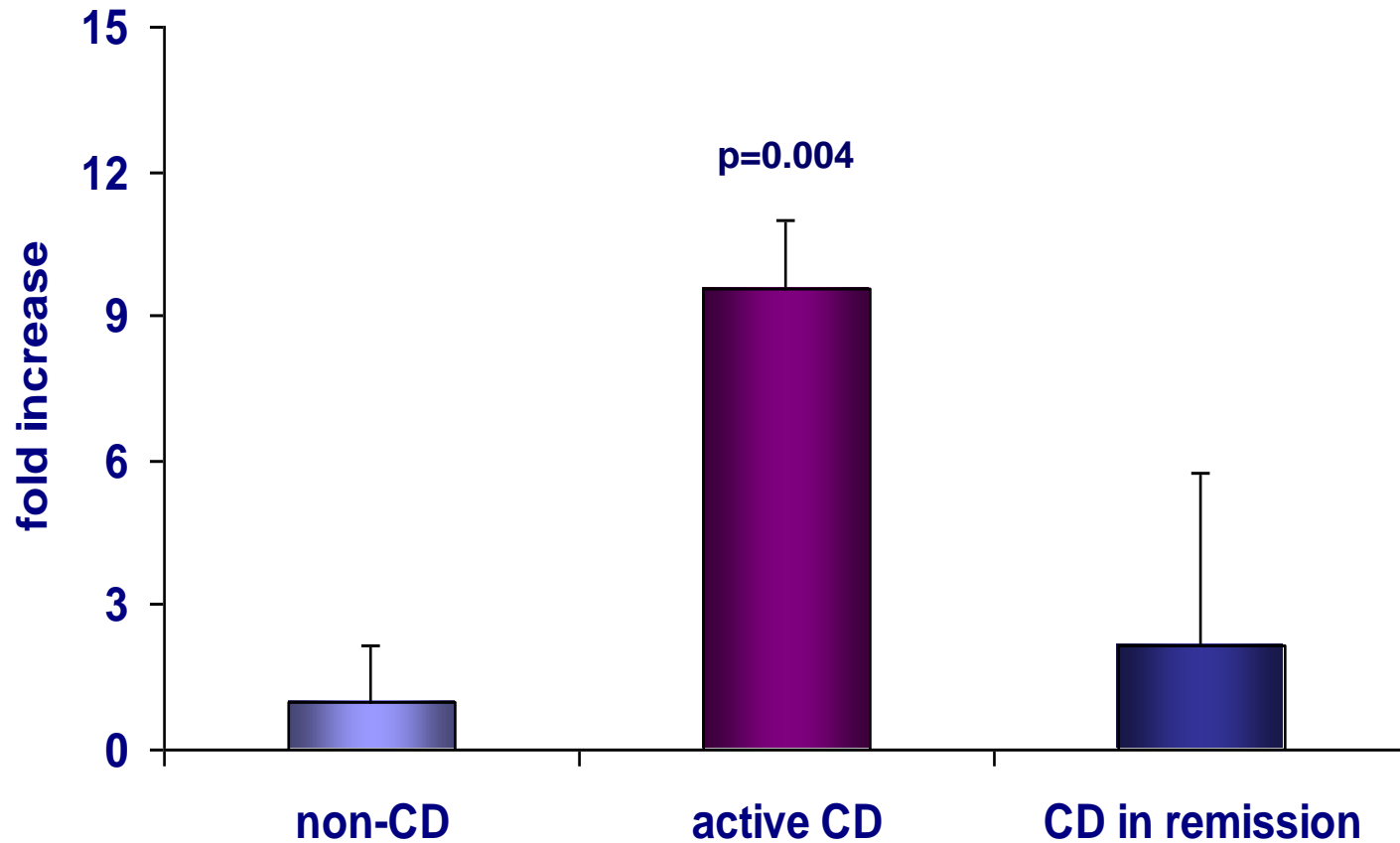


Non-celiac

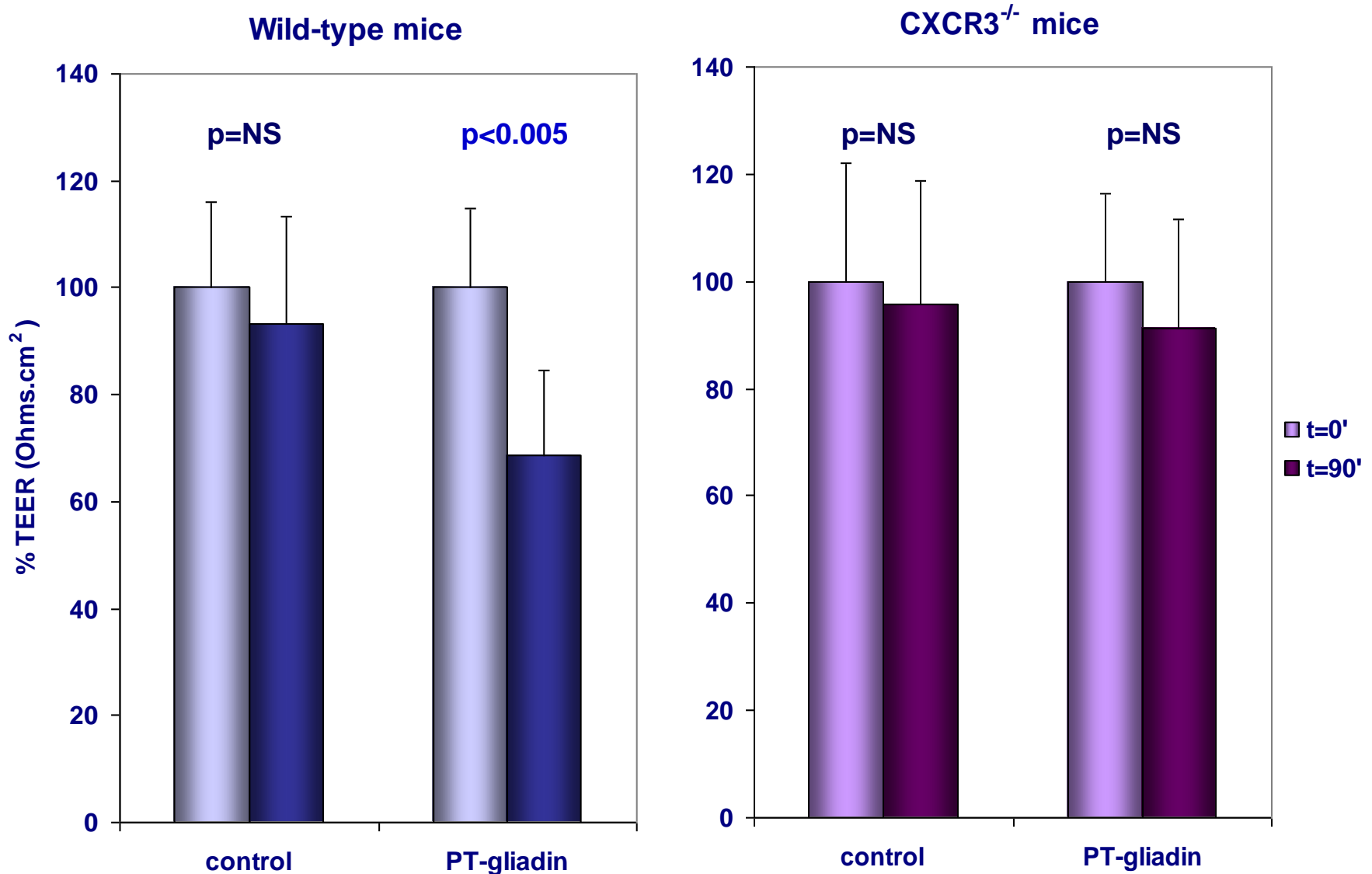




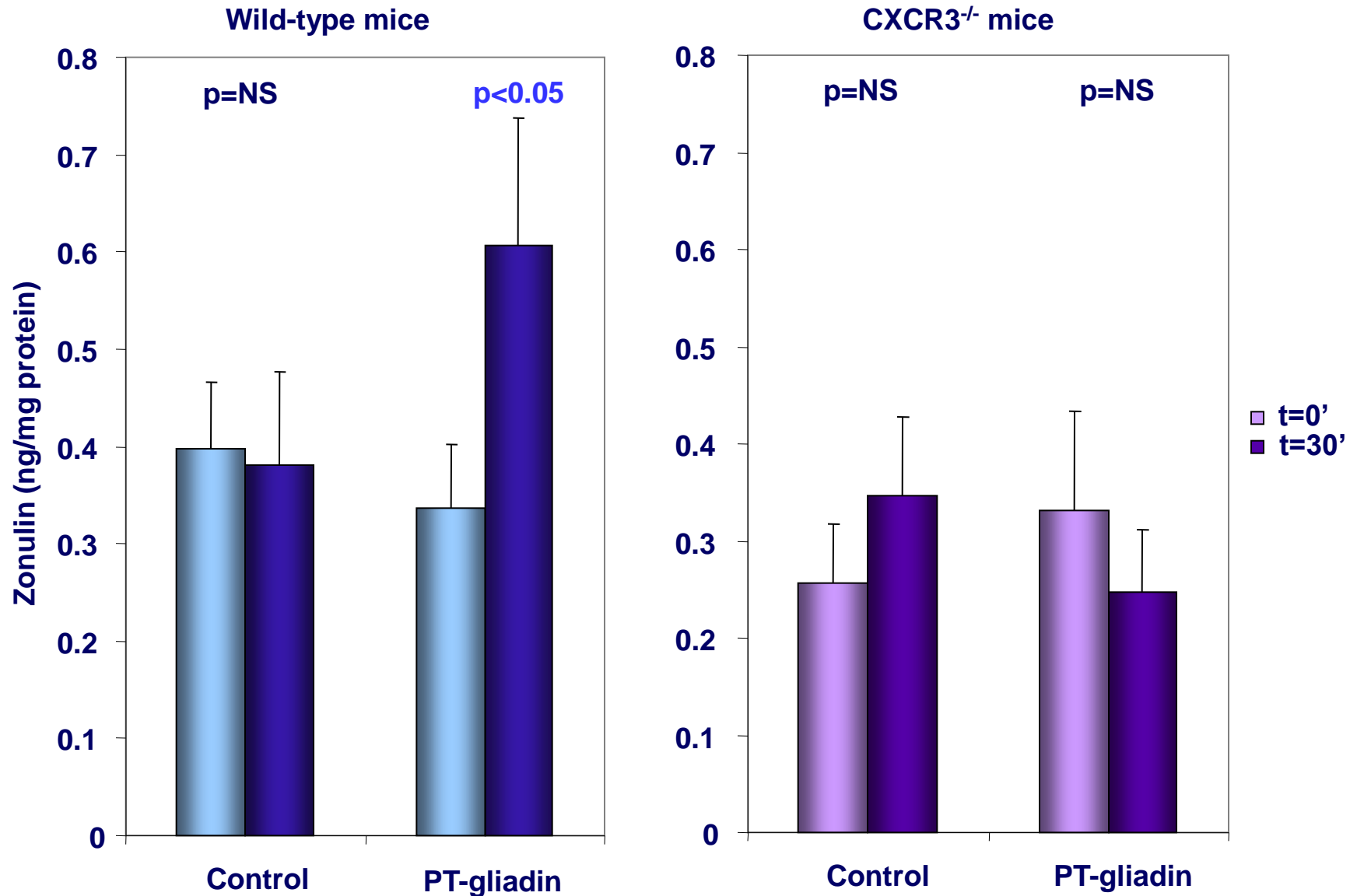
# Intestinal CXCR3 expression is elevated in active CD and returns to baseline during GFD



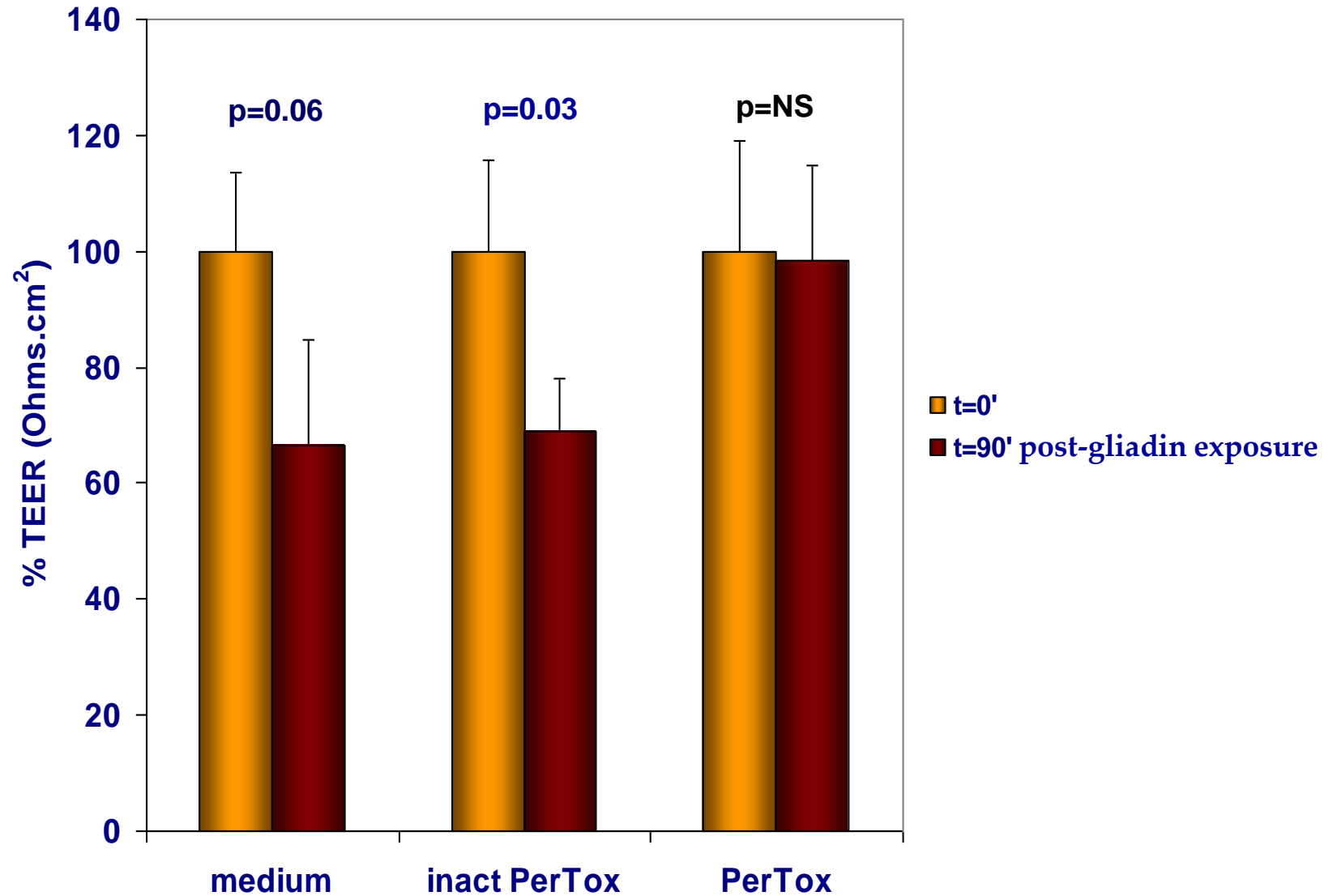
# PT-gliadin induces increased permeability in wild-type-, but not in CXCR3<sup>-/-</sup>, intestinal segments



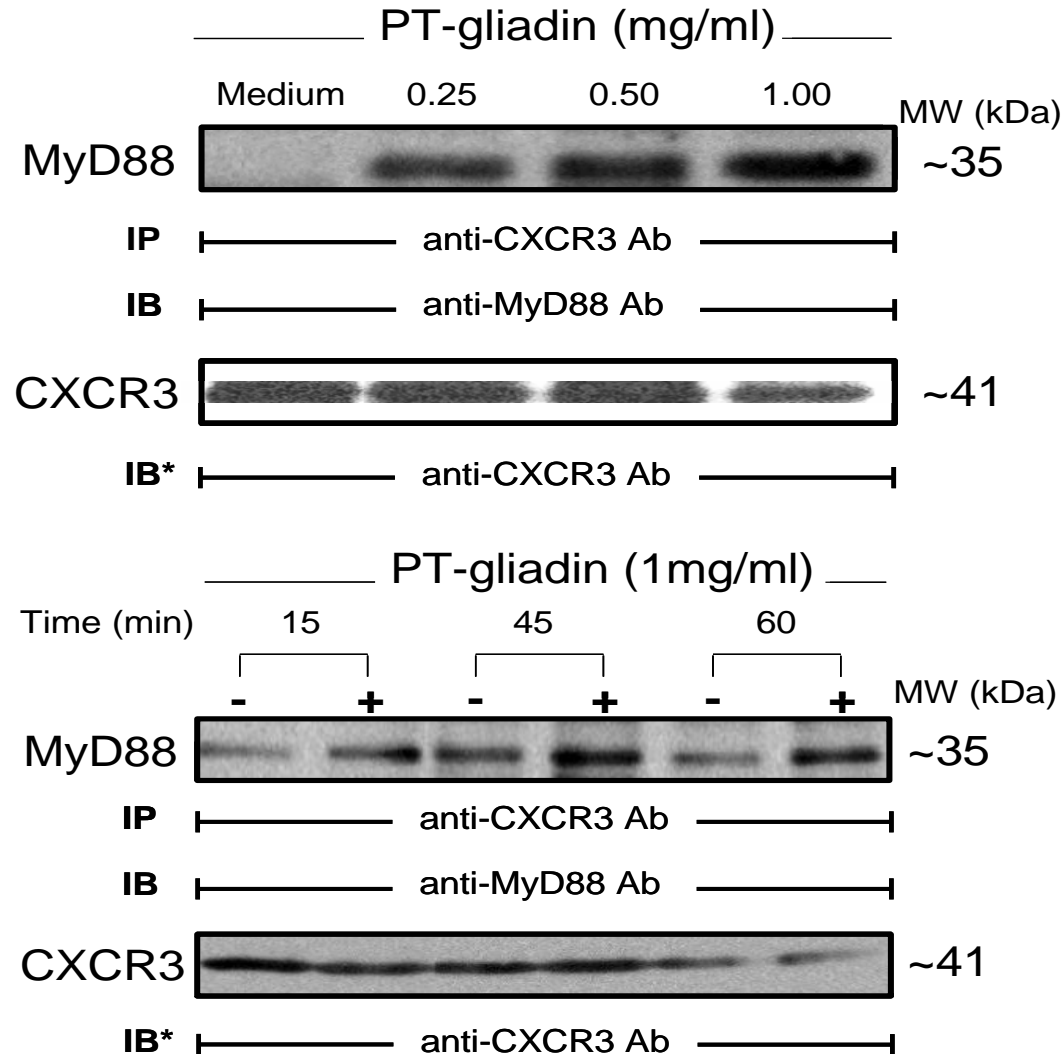
# PT-gliadin induces increased zonulin release in wild-type-, but not in CXCR3<sup>-/-</sup>, intestinal segments



# Pre-incubation with *Pertussis toxin* prevents PT-gliadin induced increase of intestinal permeability



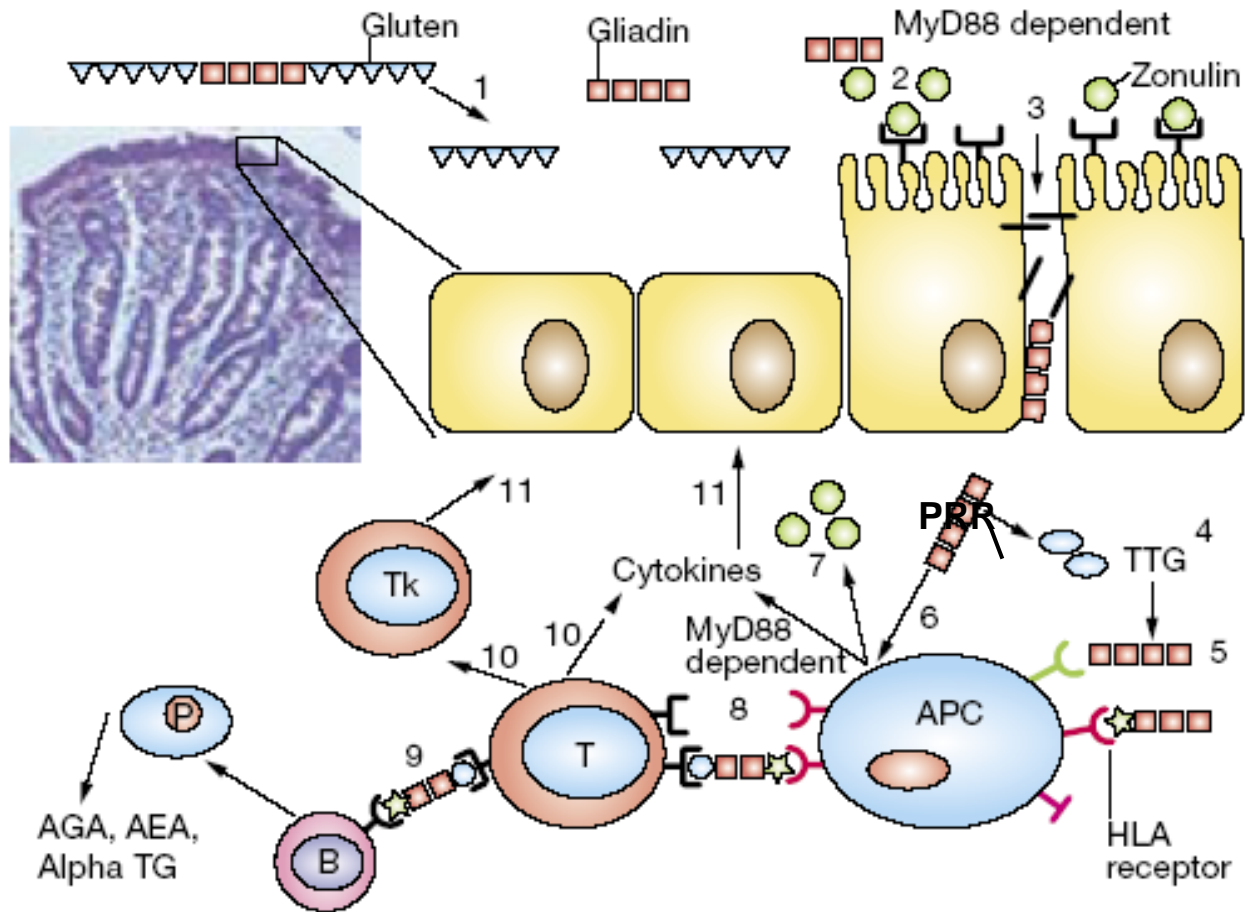
# Physical association of CXCR3 and MyD88 after PT-gliadin Binding Dose- and Time-dependent



# Conclusions

- We have shown colocalization of CXCR3 and gliadin
- CXCR3 was expressed at intestinal epithelial level, and staining showed an apical localization
- CXCR3 gene expression in biopsy specimens was enhanced in active CD compared to controls, and this expression seemed to return to base-line with GFD
- In functional studies, binding of gliadin to CXCR3 induced enhanced zonulin release and increased intestinal permeability, an early event in CD pathogenesis
- The effect of gliadin on tight junction disassembly appeared to be MyD88-dependent

# Hypothesis



- In genetically predisposed individuals binding of gliadin to CXCR3 is the first step leading to tight junction disruption and subsequent increase in intestinal permeability.
- This allows the passage of gliadin into the intestinal mucosa leading to a cascade of immune events that eventually lead to autoimmunity.